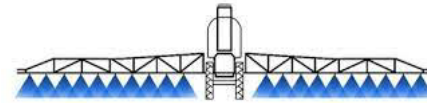


# Biopesticides for disease and insect pest management

Dave Chandler<sup>1</sup>, Elysia Bartel<sup>2</sup>, Jude Bennison<sup>2</sup>, Toby Bruce<sup>3</sup>, Luke Cartlidge<sup>4</sup>, Anca Covaci<sup>3</sup>, Clare Butler Ellis<sup>5</sup>, Elaine Fitches<sup>4</sup>, Andrew Gladman<sup>1,2</sup>, Roma Gwynn<sup>6</sup>, Rob Jacobson<sup>7</sup>, William Kirk<sup>3</sup>, Simon Leather<sup>8</sup>, Tom Pope<sup>8</sup>, Gill Prince<sup>1</sup>, Joe Roberts<sup>8</sup>, Ari Sadanandom<sup>4</sup>, Peter Seymour<sup>2</sup>, Dave Skirvin<sup>2</sup>, Graham Teakle<sup>1</sup>, Erika Wedgwood<sup>2</sup>, Sacha White<sup>2</sup>



Silsoe Spray Applications Unit



<sup>1</sup>Warwick Uni.; <sup>2</sup>ADAS; <sup>3</sup>Keele Uni; <sup>4</sup>Durham Uni; <sup>5</sup>Silsoe Spray Applications Unit, Silsoe UK; <sup>6</sup>Rationale Biopesticides; <sup>7</sup>Rob Jacobson Consultancy; <sup>8</sup>Harper Adams Uni.



SARIC • SUSTAINABLE AGRICULTURE RESEARCH & INNOVATION CLUB

# background



IBMA Global

- We need durable, sustainable IPM systems (within IFM) for all crops.
  - EU Farm 2 Fork, 50% pesticide reduction, promoting IPM.
  - COP15, 50% reduction in harmful effects of pesticides.
- Over reliance on conventional pesticides.
- We lack IPM tools.
- Many IPM tools are not being used to their full potential.
- We lack a holistic science of IPM.
- Regulatory, political, economic & technical barriers too!



PART 1: DEVELOPING  
MANAGEMENT TOOLS  
FOR BIOPESTICIDES.



PART 2: UNDERSTANDING  
INTERACTIONS IN IPM – THE  
CENTRAL ROLE OF PLANT  
GENOTYPE.

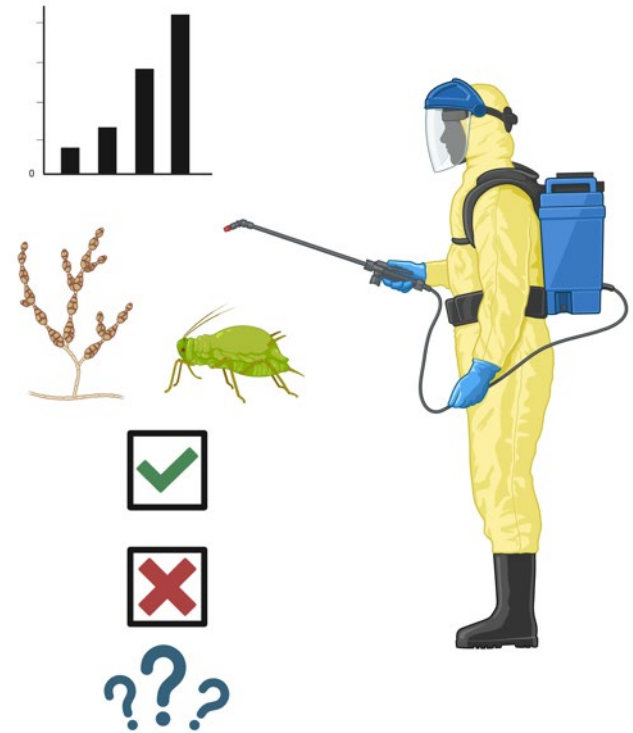
# (1) Getting the best out of biopesticides / bioprotectants

- AHDB AMBER project (Warwick, ADAS, Silsoe, consultants).
- **Application & Management of Biopesticides for Efficacy & Reliability** – PE, PO, HNS crops.
- Biopesticides – living micro-organisms & natural products.
- Develop generic management tools and practices to improve performance.



Biopesticides work well if used carefully in IPM: bioinsecticides 2<sup>nd</sup> line of defence to predators / parasitoids; biofungicides alternate with conventional fungicides

- Increasing number of products on the market.
- Only licensed products can be used: these products must show efficacy in registration trials.
- In growers' hands – a mixed picture:
- Some work well.
- Others are poor / inconsistent.
- Improve management practices: but exact reasons for suboptimal performance are often unclear.



# We observed how growers used commercial microbial biopesticide products at crop scale.

## Benchmarking trials

- Natural P&D outbreaks.
- Followed best practice guidelines.
- Compared to standard treatment if possible.

5 licensed fungal BCA products



6 growers  
(1 biopesticide, 1 P/D)



7 crops (pepper,  
cucumber, 5 ornamentals)



3 pests (aphids, thrips,  
whitefly)



3 diseases (mildew,  
botrytis, root rots)

## Detailed information on biopesticide & grower performance

*Product storage & viability; spray application; deposition on the crop; persistence; P/D control; environmental conditions; non-target effects; phytotoxicity.*



# We observed how growers used commercial microbial biopesticide products at crop scale.

- Efficacy varied: some better than conventional pesticide; others did not work at all.
- Labels & guidance hard to follow. **Lack of accessible information:** (effective dose, persistence, environmental conditions, application etc.).
- **Precision spray application needed:**
- Spray equipment not fit for purpose (1 exception).
- Lack of evidence to optimize spray application (e.g. water volumes too high).
- Not 'winning the numbers game': deliver effective dose, right place, right time.
- Record efficacy data in a consistent way to enable meaningful analysis.

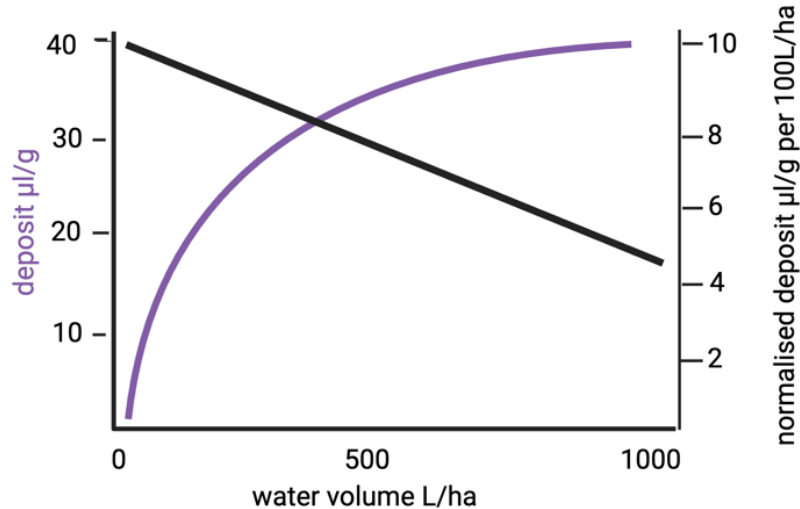


# How can we move towards precision application?

- Optimize spraying (water volume).
- Develop models to test different application strategies in silico.
- Understand how the biopesticide 'behaves' (biofungicides persistence).
- Better data recording templates (pool grower information).
- Lots of knowledge exchange.



Spray application: amount deposited is sensitive to water volume even at constant dose



Silsoe track sprayer  
experiments using tracer  
dyes (herbs, ornamentals,  
tomato)

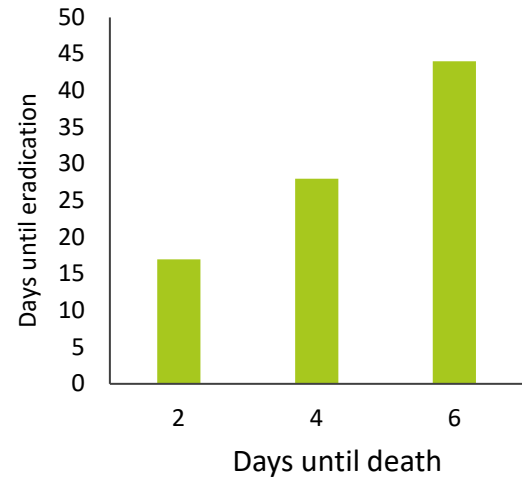


- Label recs for hort crops are up to 1500 L / ha.
- Inefficient, wasteful.
- Nominal constant dose application, amount of a.i. deposited on plants is actually sensitive to water volume.
- Lower volumes deposit more a.i. / leaf area. Spraying is faster, less waste.
- Increasing the concentration of a.i. on leaves increases efficacy.
- Water needed for activation?
- Biopesticides have an optimum water volume for efficacy - but companies don't know what this is!

# Precision application: a boxcar model of pest development to inform biopesticide use strategy

- Whitefly, aphids.
- Tracks the maturation of individuals to next life stage, reproduction & lifespan.
- Add in biopesticide mortality.
- **Test out biopesticide strategies in silico** (persistence, infection efficacy, speed of kill, frequency, pest population size).

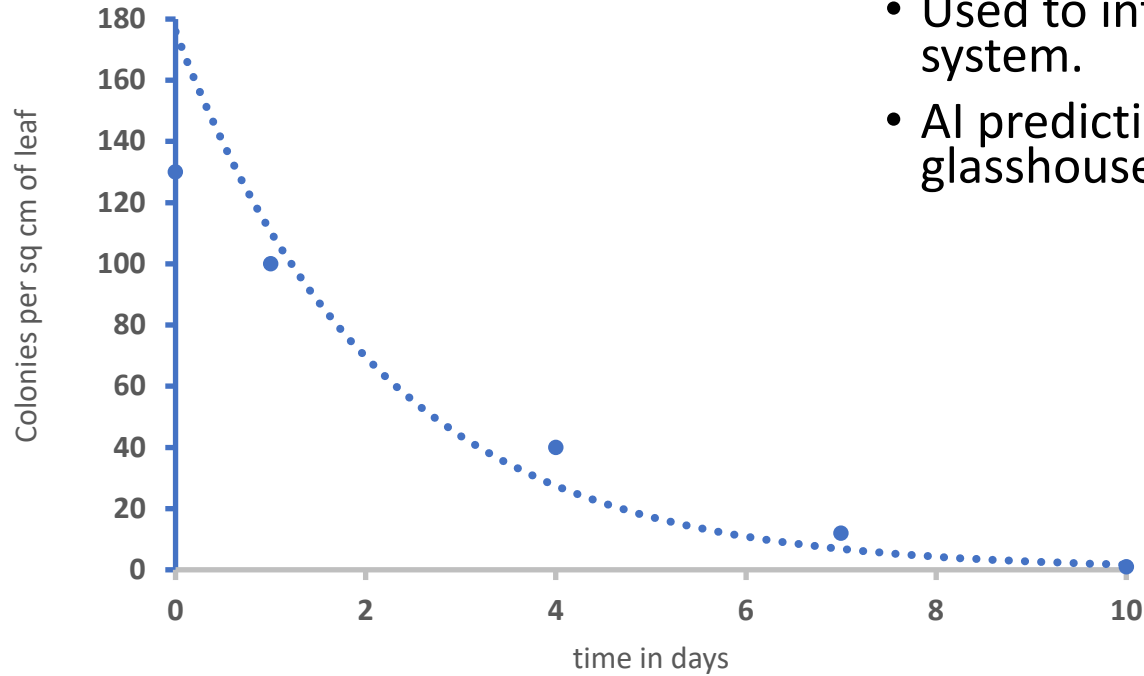
Effect of EPF speed of kill on whitefly growth (using dummy data)



- Initial population size = 1000 adults.
- Sprayed on day 7, 14, 21, 28 and 35.
- All stages affected.
- Infection efficacy = 90%.
- Persistence = 5 days.



# Powdery mildew control: Short survival on leaves of the obligate mycoparasite *Ampelomyces* in absence of its host



- Used to inform a smart decision support system.
- AI prediction of PM risk period from glasshouse environmental data.

Some growers assumed *Ampelomyces* was a preventative



Narrow use window

# Improving access to information

- Grower articles, talks, webinars - 9 crop sectors.
- Application workshops (> 100 growers / agronomists).
- Literature reviews to summarize how conditions affect performance (biofungicides).
- New data recording template for growers.



# Biopesticides: 'winning the numbers game'.

Biopesticides need precision application, based on detailed understanding of their mode of action.

Strategic thinking & funding for UK crop protection.

Other areas that require investment:

Formulation.

Mass production.

Use in IPM systems.



## (2) A holistic science of IPM – interactions in the system

Biological crop protection:  
a 'slow down / speed up' strategy for  
aphid management on brassica

**SARIC** • SUSTAINABLE AGRICULTURE  
RESEARCH & INNOVATION CLUB

Creative commons



Harper Adams  
University







## Aphids: Combining durable crop resistance with biocontrols

- *Myzus persicae* & *Brevicoryne brassicae* on Brassica crops.
- Partial (durable) host plant resistance.
- Conservation control with parasitoids.
- Plant defence activator cis-jasmone.
- Entomopathogenic fungi as a biopesticide.

5

Interactions in the system

3

Alter activity of aphids & their parasitoids with plant volatile cis-jasmone

2

Gene expression responses

Targeted plant breeding

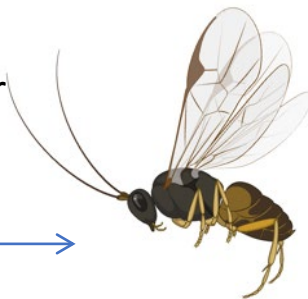
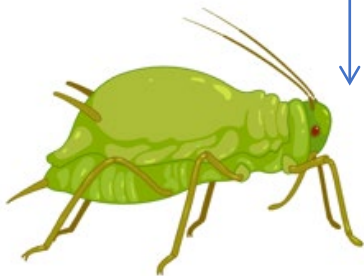
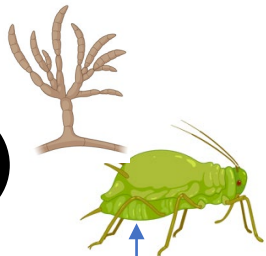
1

Identify partial resistance in Brassica

4

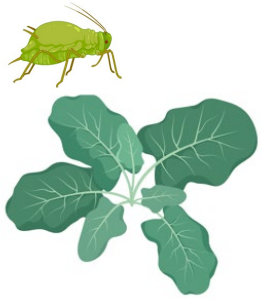
EPF against aphid nymphs

Slow aphid development?



# Key findings: putting crop genotype at the centre of IPM

1

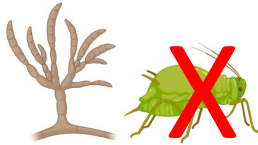


2

SA

Resistance in *B. oleracea* is associated with upregulation of the SA pathway

Partial resistance in *B. oleracea*, *B. cretica*, *B. napus* – includes inbred lines

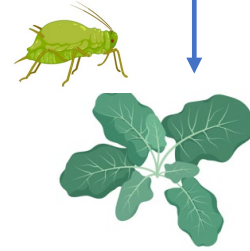


5



Partially resistant plants make aphid nymphs susceptible to fungal biopesticides

cis-jasmone



3

Cis-jasmone activates parasitoid activity & affects aphid behaviour depending on plant genotype.

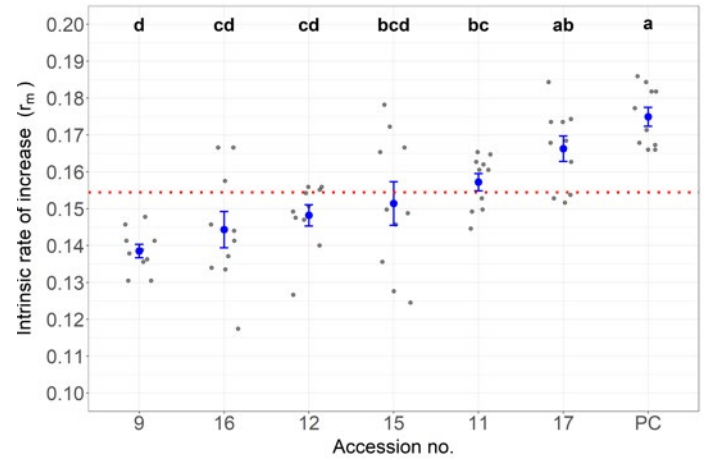
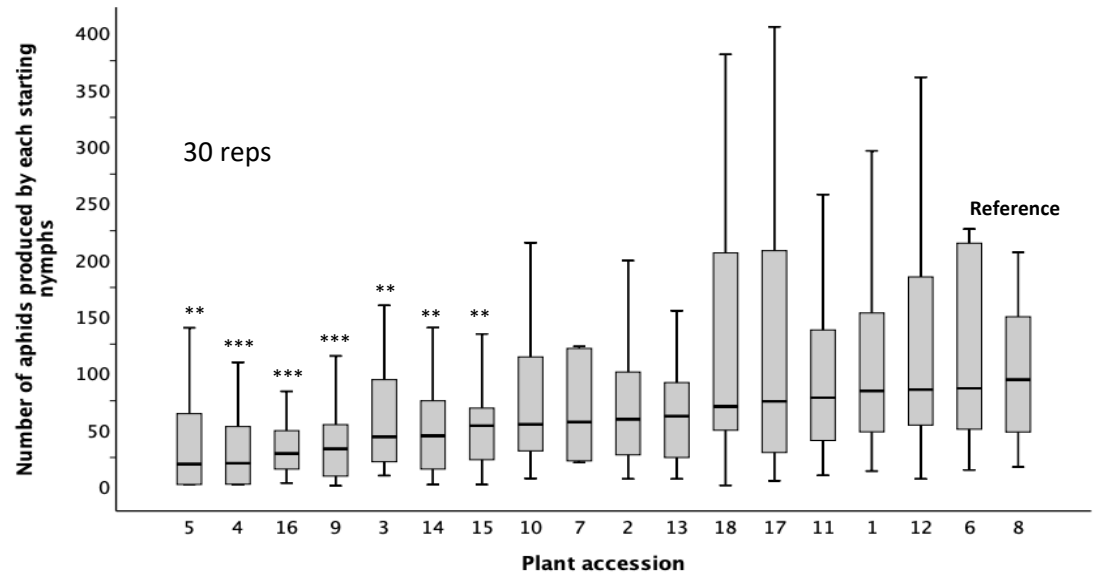
4



Natural populations of parasitoids are active in most field sites, giving opportunities to use them in conservation biological control.

1

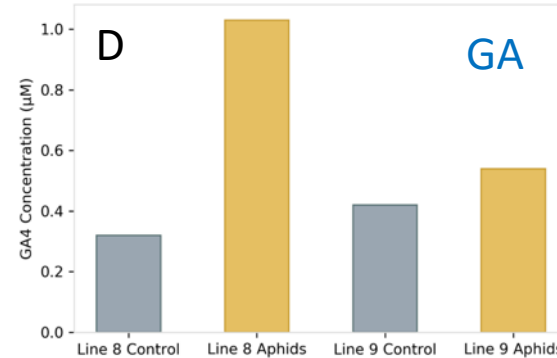
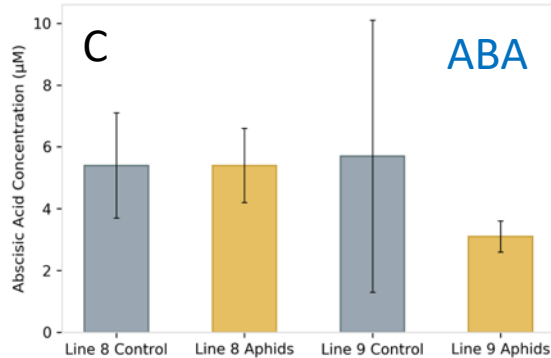
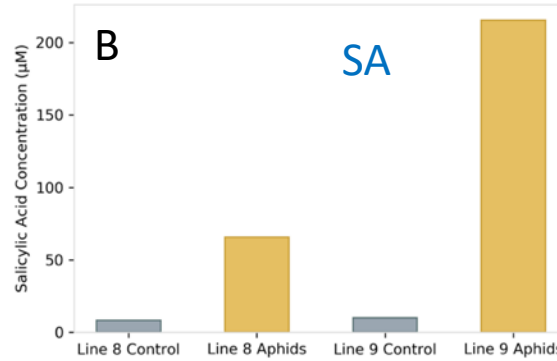
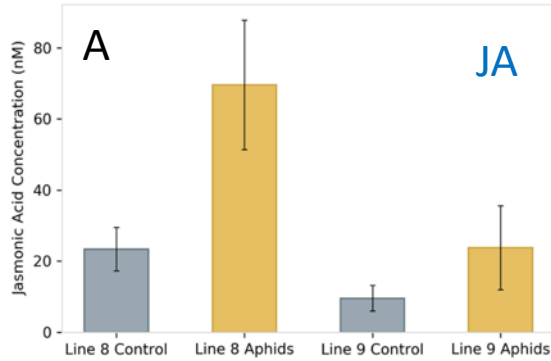
Partial resistance to *Myzus* & *Brevicoryne* identified in vegetable brassicas (& *B. napus*). Confirmed in field cage experiments. Associated with reduced intrinsic rate of increase.



**Resistant -> Susceptible**  
**16, 9, 15, 11, 12, 17**

Backed up by field cage studies

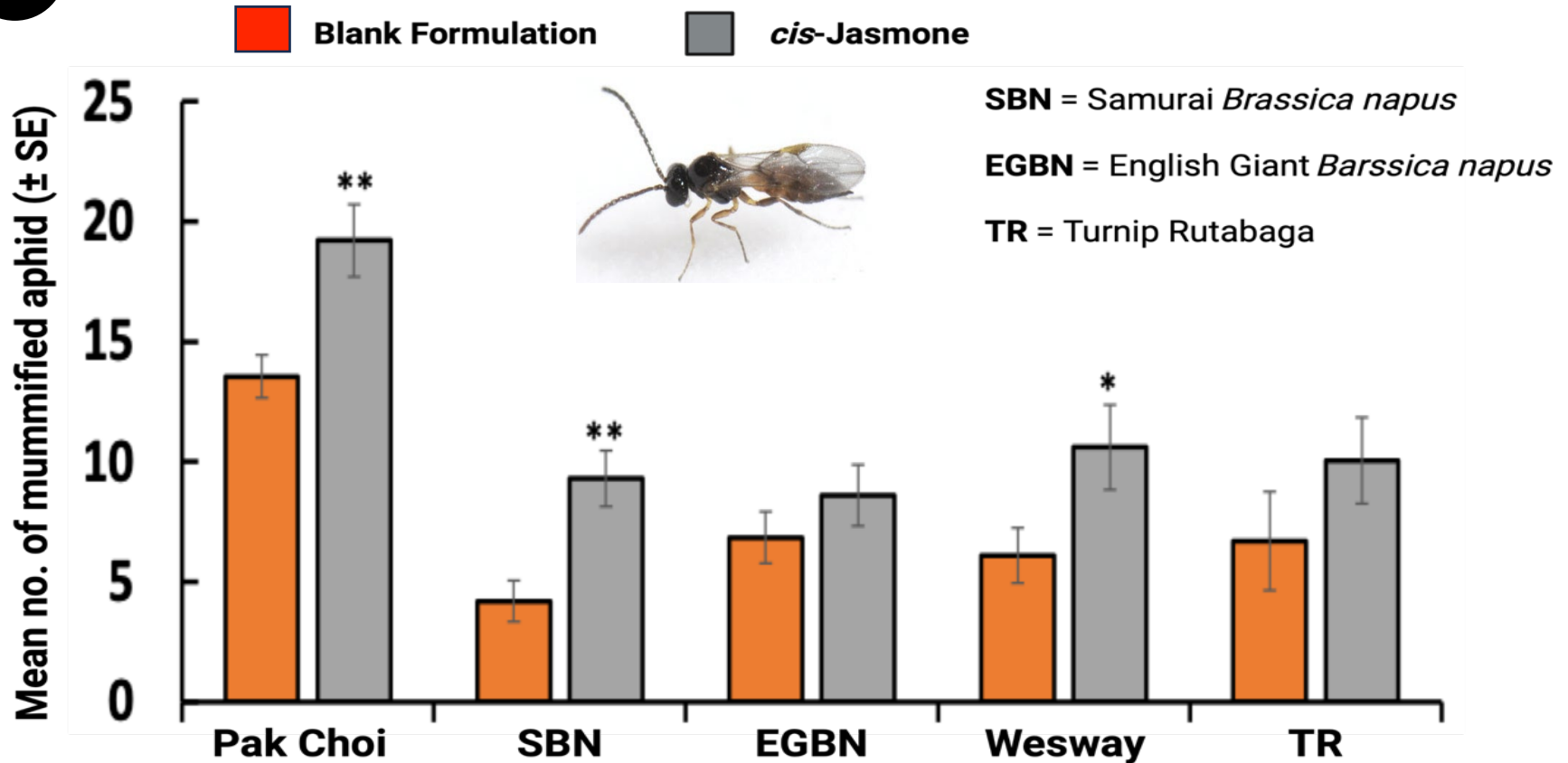
## Partial resistance linked to the SA signalling pathway



- RNA seq: more DE genes in line 8 >> 9 at 2h & 6 h.
- Reversed at 24 h.

- Concentrations of **A**: Jasmonic acid, **B**: Salicylic acid, **C**: Abscisic acid and **D**: GA4 from mass spec. leaf tissue of *Brassica oleracea*.
- 8= susceptible.
- 9 = resistant.
- 20 *Myzus* in clip cage for 24 h.
- Previous work identifies JA as the defence pathway – but have not worked with R vs S plants.

# *cis*-Jasmone treatment increased parasitism (lab)

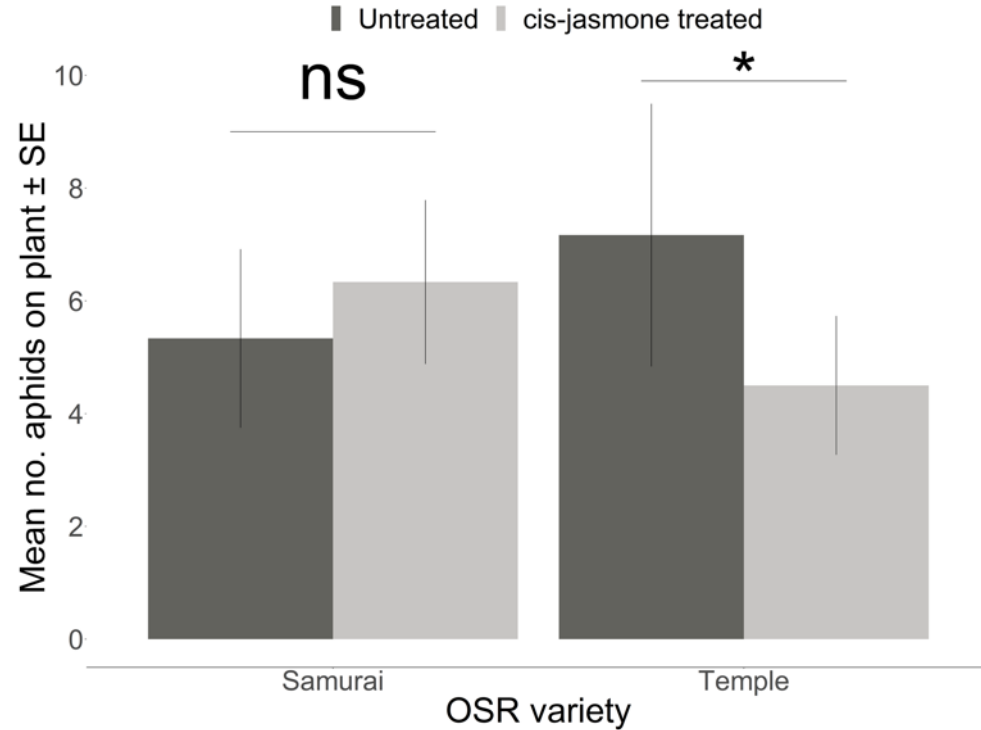
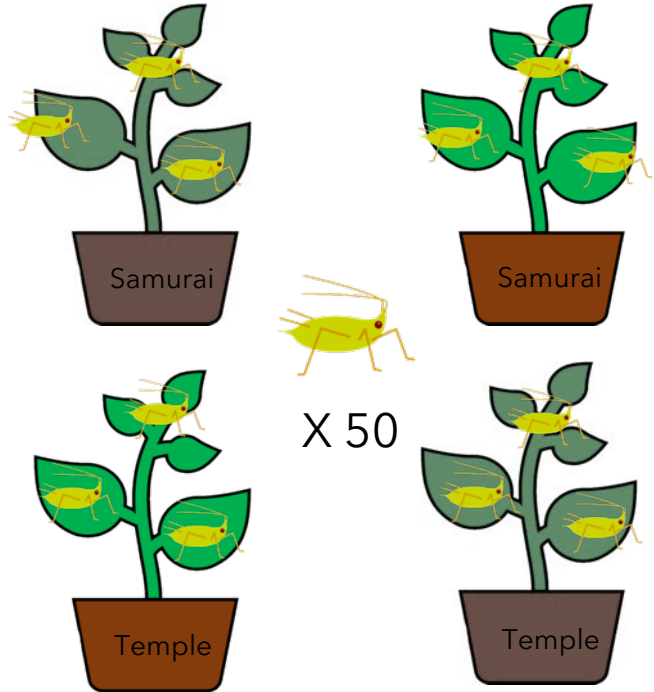




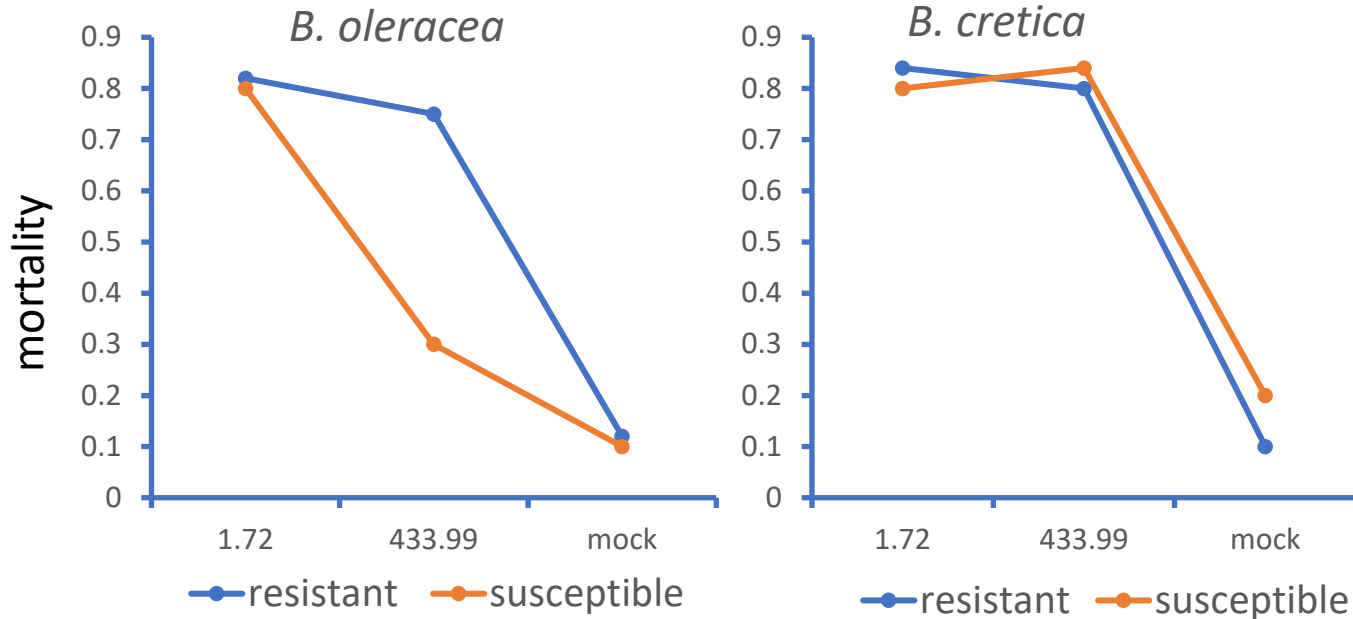
# Field trial

*Aphid landing behaviour*

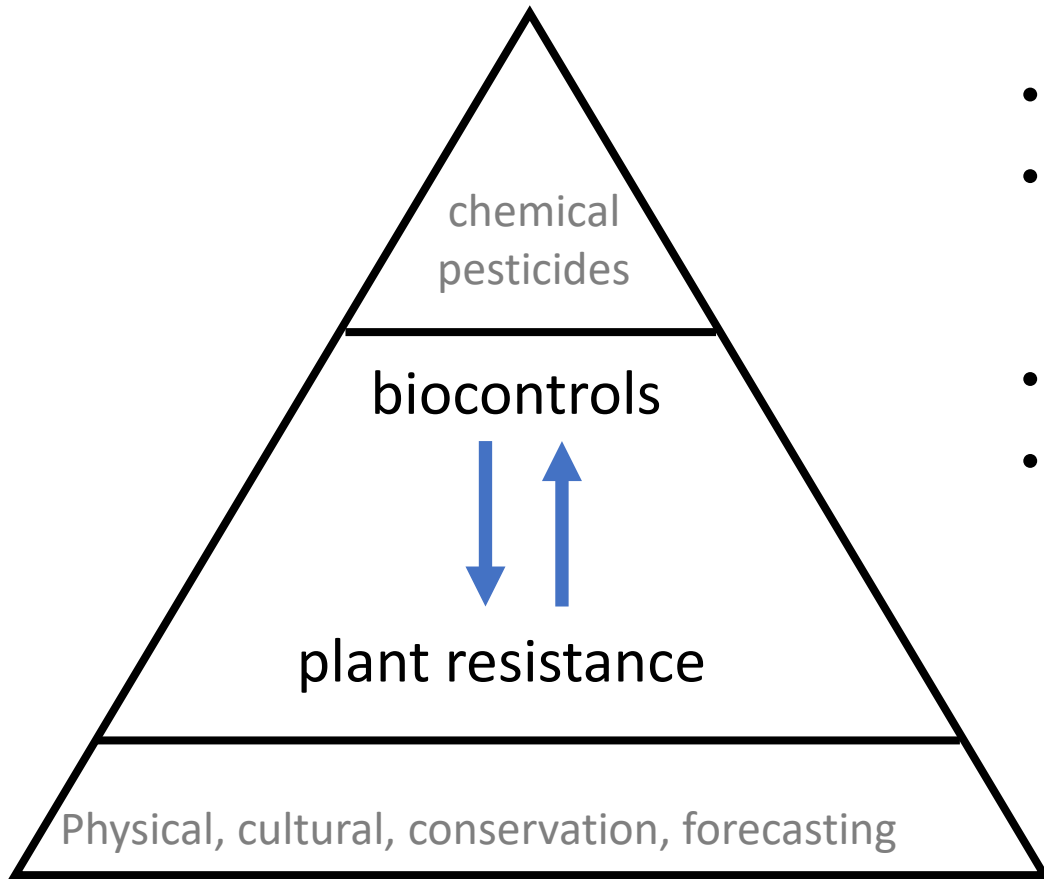
24 hrs



# Brassica genotype makes aphid nymphs susceptible to fungal pathogen



- Strain 1.72 kills nymphs of *Myzus* & *Brevicoryne*.
- But strain 1.72 is no longer available as a commercial biopesticide.
- 433.99 is available as a product but does not normally kill nymphs.
- Partially resistant *B. oleracea* – and *B. cretica* – make nymphs susceptible.



- Future IPM:
- Multi-trait crop improvements that work in synergy with biocontrols / bioprotectants.
- Holistic IPM science.
- We need a strategic plan for IPM.

Thank you

